

Microglia influence host defense, disease, and repair following murine coronavirus infection of the CNS

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Day 7 post-infection - Brain Introduction **Microglia depletion increases demyelination** Day 21 Brain 🔵 Day 7 - Control 😑 Day 7 – PLX5622 **Day 14** Background: PLX562 • Mice infected with the neurotropic JHM strain of mouse hepatitis virus (JHMV) suffer from acute encephalomyelitis and chronic demyelination within the central nervous system (CNS). • Immunologic control of viral replication within the CNS is complex, B cells Monocytes involving both the innate and adaptive immune response. Control PLX5622 Control PLX5622 • Neuropathology is mediated by local glial responses working in Day 14 - Control Day 14 – PLX5622 Microglia Neuts Mac 1 Mac 2 combination with components of the innate and adaptive responses. **Goal**: To understand the functional contributions of microglia in host Xcr1+ DCs defense and demyelination in response to JHMV infection of the CNS Xcr1+DCs Ccl22+ DCs Nox2+ DCs pDCs Approach: • Treated mice with PLX5622, an inhibitor of colony stimulating factor 1 receptor (CSF1R), that efficiently depletes microglia. Figure 6: (A) Representative images of H&E/LFB-stained spinal cord sections showing

• Conducted single cell RNA sequencing (scRNASeq) on CD45+ cells

Figure 2: (A) Unsupervised clustering analysis of cells from brains at day 7 p.i. following scRNAseq. (B) DotPlot representing percentage of cells from control (blue) versus PLX5622 (orange) within a cluster

- isolated from the CNS of PLX5622 and control-treated mice at days 7 and 14 p.i.
- Evaluated spinal cord demyelination at day 14 and 21 p.i. and remyelination at day 21 p.i.



PLX5622 Treatment:



(pooled brains from 5-6 mice/experimental group).

Depletion of microglia dampens macrophage APC activity and CD4⁺ T cell responses



increased demyelination (dashed lines) in JHMV-infected mice treated with PLX5622 vs controls at 14 d.p.i. (B) Quantification of demyelination reveals a significant increase in PLX5622-treated mice at days 14 and 21 p.i. (min. of 12 mice/group). (C) Expression of Apoe, Spp1(Osteopontin), and Trem2 in spinal cords of PLX5622-treated (green) versus controls (grey) in all macrophages at 14 d.p.i. *p≤ 0.05; **p≤ 0.01; ****p≤ 0.0001.

Microglial depletion reduces capacity to remyelinate







Figure 3: Brains of control (black, blue) and PLX5622-treated (grey, orange) mice at 7 d.p.i. (A) Quantification of flow cytometric data of infiltrating CD4 and CD8 T cells (n=6/group). Box plots comparing expression of activation markers Cd44, Il2ra, Il2rb, and Tbx21 (T-bet) in CD4 T cells (B), effector markers Prf1 (Perforin), *Pdcd1* (Programmed cell death 1), and *Gzmb* in CD8 T cells (**C**), and MHC class II genes and co-stimulatory molecule Cd86 in macrophages (D). Plots show interquartile range, median value (bold horizontal bar) and average expression per sample (red dot). Used Wilcoxon test, *p≤ 0.05; ** p≤ 0.01; ****p≤ 0.0001.

Day 14 and Day 21 post-infection – Spinal Cord





Figure 5: (A) EM images (1200X) from PLX5622 and control spinal cords showing normal myelinated axons (white arrowheads), demyelinated axons (red arrows) and remyelinated axons (blue arrows) at day 21 p.i. (B) Calculation of g-ratio (axon diameter/total fiber diameter) of PLX5622 and control. Box plots comparing expression of transcripts of remyelination-associated markers *Cst7* (Cystatin F), *Igf1* (Insulin-like growth factor 1), and *LpI* (Lipoprotein lipase) in merged microglia (C) and macrophage (D) populations in spinal cords of control (green) and PLX5622-treated (grey) mice. $p \le 0.05$; $p \le 0.0001$.

Perspectives

Day 7 p.i. – Brains

Mac 2

/licroglia (MG

Nox2+ DCs

- Microglia aid in control of viral replication
- Depletion of microglia results in an inability to prime a Th1 response in CD4⁺ T cells

Day 14 and 21 p.i. – Spinal Cords

Microglia potentially play a protective role by restricting white matter pathology and contributing to remyelination

Acknowledgements

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Figure 4: (A) Unsupervised clustering analysis of cells from spinal cords at 14 d.p.i. after scRNAseq. (B)

DotPlot representing percentage of cells from control (green) and PLX5622-treated (grey) mice within a

cluster (pooled spinal cords from 6-7 mice/experimental group).



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